# LC-MS/MS Analysis of 20 Underivatized Amino Acids on Supel<sup>™</sup> Carbon LC column

## Introduction

Amino acids are the building blocks of proteins and peptides in biological systems. Amino acids have also been used as supplements to support health and as indicators for certain diseases. The challenge with analyzing amino acids is due to the wideranging polarities spanning across all 20 amino acids (**Figure 1**). Due to this complexity, methods in the past have relied on derivatization of the amino acids; however, derivatization can lead to further complexity due to the presence of derivatized/underivatized amino acids and interferences from the derivatizing reagent itself. This application outlines an LC-MS/ MS method for analyzing all 20 amino acids, without derivatization, utilizing the Supel<sup>™</sup> Carbon LC column. All 20 amino acids are retained on the column, with good peak shape.

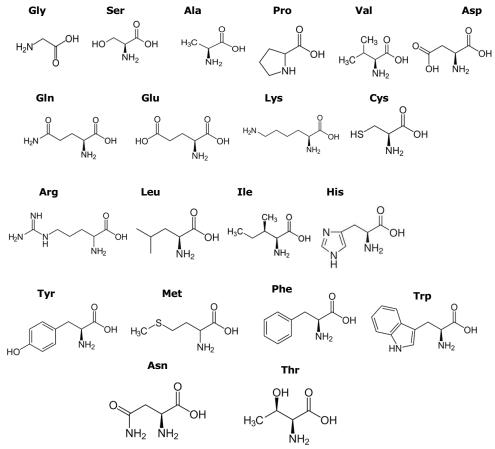


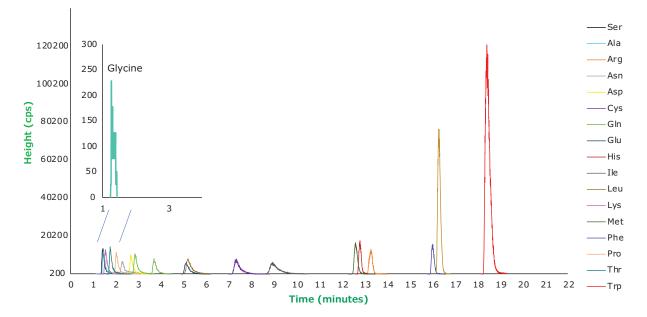
Figure 1: Structures and three letter codes of amino acids.



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# **Results/Conclusion**

**Figure 2** displays the MS spectral results for the analysis of 20 underivatized amino acids while **Table 1** displays the optimized MS conditions for the separation and **Table 2** displays the fragment ions and fragmentation parameters for the amino acids.



## Supel<sup>™</sup> Carbon LC column - 20 Underivatized Amino Acids

Figure 2: Separation of 20 underivatized amino acids by LC-MS/MS. Conditions: Column: Supel<sup>™</sup> Carbon LC, 10 cm x 2.1 mm I.D., 2.7 μm; Mobile Phase: [A] Water (0.1% (v/v) DFA); [B] Acetonitrile (0.1% (v/v) DFA); Gradient: Hold at 0% B for 7 min; 0% B to 5% B in 5 min; 5% B to 100% B in 10 min; Flow Rate: 0.2 mL/min; Column temp.: 12 °C; Detector: MSD; Injection: 1.0 μL; Sample: Amino Acid Mix, varied concentration, water (0.1% (v/v) DFA)

| Elution<br>Order | Compound      | Retention Time<br>(min) | Elution<br>Order | Compound      | Retention Time<br>(min) |
|------------------|---------------|-------------------------|------------------|---------------|-------------------------|
| 1                | Glycine       | 1.27                    | 11               | Glutamic Acid | 5.13                    |
| 2                | Serine        | 1.43                    | 12               | Leucine       | 5.18                    |
| 3                | Alanine       | 1.45                    | 13               | Cystine       | 7.34                    |
| 4                | Lysine        | 1.54                    | 14               | Isoleucine    | 8.93                    |
| 5                | Threonine     | 1.75                    | 15               | Methionine    | 12.61                   |
| 6                | Proline       | 2.03                    | 16               | Histidine     | 12.81                   |
| 7                | Asparagine    | 2.29                    | 17               | Arginine      | 13.30                   |
| 8                | Aspartic Acid | 2.65                    | 18               | Phenylalanine | 16.03                   |
| 9                | Valine        | 2.85                    | 19               | Tyrosine      | 16.29                   |
| 10               | Glutamine     | 3.69                    | 20               | Tryptophan    | 18.42                   |

#### Table 1: MS Conditions for the Separation of 20 Amino Acids

| Ion Source Type   | Turbo Spray |
|-------------------|-------------|
| Curtain Gas       | 25          |
| Ion Spray Voltage | 4000 V      |
| Temperature       | 300 °C      |
| Ion Source Gas 1  | 20          |
| Ion Source Gas 2  | 30          |
| Interface Heater  | On          |

## Table 2: MRM Fragmentation Parameters and Ions for the 20 Amino Acids

| Name          | Q1    | Q3    | DP   | EP  | CEP  | CE    | СХР |
|---------------|-------|-------|------|-----|------|-------|-----|
| Lysine        | 147.1 | 84.0  | 21.7 | 7.0 | 5.3  | 21.0  | 3.0 |
| Proline       | 116.1 | 70.1  | 22.2 | 7.9 | 6.2  | 19.1  | 4.1 |
| Aspartic Acid | 134.1 | 74.0  | 17.0 | 8.3 | 12.8 | 17.0  | 3.7 |
| Serine        | 106.1 | 60.0  | 13.0 | 4.0 | 8.8  | 15.0  | 3.1 |
| Glycine       | 76.1  | 30.0  | 7.0  | 7.0 | 4.9  | 17.0  | 5.5 |
| Alanine       | 90.1  | 44.0  | 16.0 | 6.0 | 4.9  | 16.9  | 5.6 |
| Threonine     | 120.1 | 56.0  | 19.0 | 5.3 | 7.0  | 22.0  | 3.3 |
| Asparagine    | 133.1 | 87.0  | 17.0 | 3.3 | 8.2  | 14.0  | 2.6 |
| Valine        | 118.1 | 72.0  | 14.5 | 6.0 | 7.5  | 14.0  | 3.2 |
| Glutamine     | 147.1 | 84.0  | 12.0 | 6.8 | 6.1  | 23.0  | 3.5 |
| Leucine       | 132.1 | 86.0  | 16.0 | 7.4 | 10.2 | 14.0  | 3.2 |
| Glutamic Acid | 148.1 | 84.0  | 24.0 | 6.1 | 9.0  | 20.0  | 3.0 |
| Isoleucine    | 132.1 | 86.0  | 16.0 | 7.4 | 10.2 | 14.0  | 3.2 |
| Methionine    | 150.2 | 104.0 | 16.0 | 5.5 | 11.8 | 13.6  | 3.9 |
| Histidine     | 156.1 | 110.0 | 24.0 | 3.5 | 11.0 | 19.0  | 3.7 |
| Arginine      | 175.2 | 70.0  | 25.2 | 6.7 | 10.0 | 37.7  | 3.0 |
| Phenylalanine | 166.2 | 120.2 | 18.0 | 7.0 | 9.1  | 16.8  | 3.2 |
| Tryptophan    | 205.2 | 146.2 | 19.4 | 4.8 | 17.7 | 23.0  | 4.0 |
| Tyrosine      | 182.2 | 136.2 | 19.2 | 7.7 | 6.4  | 18.04 | 4.0 |
| Cystine       | 241.2 | 120.0 | 26.0 | 7.8 | 9.8  | 26.0  | 4.1 |

This application demonstrated the effectiveness of the Supel<sup>™</sup> Carbon LC column in resolving 20 amino acids without the need for a derivatization reagent. Analysis of amino acids is typically done with speciality, silica-based columns, ion exchange columns, or normal phase columns. However, by utilizing tandem MS/ MS detection, analyses of amino acids can be accomplished on Supel<sup>™</sup> Carbon LC column with fast run times compared to most commercial approaches. Due to porous graphitic carbon's unique ability to discriminate three-dimensional differences between compounds, Leucine and Isoleucine can be resolved with exceptional resolution.

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