

# **Restriction Endonuclease Hpa I**

From Haemophilus parainfluenza

**Cat. No.** 10 380 385 001 100 units (3-10  $U/\mu l$ ) **Cat. No.** 10 567 647 001 500 units (3-10  $U/\mu l$ )



**Version 15** Content version: June 2017

Store at -15 to -25°C

Stability/Storage

The undiluted enzyme solution is stable when stored at -15 to  $-25^{\circ}$ C until the control date printed on the label. Do not store below  $-25^{\circ}$ C to avoid freezing.

Sequence specificity

*Hpa* I recognizes the sequence GTT/AAC and generates fragments with blunt ends (1).

Compatible ends

The enzyme generates compatible ends to any blunt end

Isoschizomers

Hpa I is not known to have isoschizomers.

Methylation sensitivity

Hpa I is inhibited by the presence of 6-methyladenine (\*) and 5-hydroxymethylcytosine. 5-Methylcytosine does not influence the cleavage (°)

Storage buffer

buffer, 10x

System

20 mM Tris-HCl, 50 mM KCl, 0.5 mM EDTA, 5 mM 2-Mercaptoethanol, 50% Glycerol (v/v), 0.01% Polydocanol, pH approx. 7.5 (at 4°C).

Suppl. Incubation

330 mM Tris-acetate, 660 mM K-acetate, 100 mM Mg-acetate, 5 mM Dithiothreitol, pH 7.9 (at 37°C), (= SuRE/Cut Buffer **A)** 

Activity in SuRE/Cut Buffer

Bold face printed buffer indicates the recommended buffer for optimal activity:

Α	В	L	M	Н
100%	25-50%	25-50%	50-75%	25-50%

Incubation temperature

ation 37°C

**Unit definition** 

One unit is the enzyme activity that completely cleaves 1 $\mu$ g  $\lambda$ DNA in 1 h **at 37°C** in a total volume of 25  $\mu$ l SuRE/Cut buffer **A.** 

Typical experiment

Component	Final concentration
DNA	1 μg
10 × SuRE/Cut Buffer <b>A</b>	2.5 μl
Repurified water	Up to a total volume of 25 μl
Restriction enzyme	1 unit

Incubate at 37°C for 1 h.

**Heat inactivation** 

 $\mathit{Hpa}\,I$  cannot be heat inactivated by heating to 65 °C for 15 min.

Number of cleavage sites on different DNAs (2):

λ	Ad2	SV40	Φ X174	M13mp7	pBR322	pBR328	pUC18
14	6	4	3	0	0	0	0

Activity in PCR buffer

Relative activity in PCR mix (Taq DNA Polymerase buffer) is 100%. The PCR mix contained  $\lambda$  target DNA, primers, 10 mM Tris-HCl (pH 8.3, 20°C), 50 mM KCl, 1.5 mM MgCl $_2$ , 200  $\mu$ M dNTPs, 2.5 U Taq DNA polymerase. The mix was subjected to 25 amplification cycles.

Ligation and recutting assay

*Hpa* I fragments obtained by complete digestion of 1 μg λDNA are ligated with 1 U T4-DNA ligase (Cat. No. 10 481 220 001) in a volume of 10 μl by incubation for 16 h at 4°C in 66 mM Tris-HCl, 5 mM MgCl<sub>2</sub>, 5 mM Dithiothreitol, 1 mM ATP, pH 7.5 (at 20°C) resulting in >80 % recovery of 1 μg λDNA × *Hpa* I fragments. Subsequent re-cutting with *Hpa* I yields > 95% of the typical pattern of λDNA × *Hpa* I fragments

**Troubleshooting** 

A critical component is the DNA substrate. Many compounds used in the isolation of DNA such as phenol, chloroform, ethanol, SDS, high levels of NaCl, metal ions (e.g., Hg²+, Mn²+) inhibit or alter recognition specificity of many restriction enzymes. Such compounds should be removed by ethanol precipitation followed by drying, before the DNA is added to the restriction digest reaction. Appropriate mixing of the enzyme is recommended.

**Quality control** 

Lot-specific certificates of analysis are available at www.lifescience.roche.com/certificates.

Absence of unspecific endonuclease activities 1  $\mu$ g  $\lambda$ DNA is incubated for 16 h in 50  $\mu$ l SuRE/Cut buffer A with excess of *Hpa* l. The number of enzyme units which do not change the enzymespecific pattern is stated in the certificate of analysis.

Absence of exonuclease activity

Approx. 5  $\mu$ g [ $^3$ H] labeled calf thymus DNA are incubated with 3  $\mu$ l Hpa I for 4 h at 37°C in a total volume of 100  $\mu$ l 50 mM Tris-HCl, 10 mM MgCl $_2$ , 1 mM Dithioerythritol, pH approx. 75. Under these conditions, no release of radioactivity is detectable, as stated in the certificate of analysis.

# References

- 1 Garfin, D. E. & Goodman, H. M. (1974) Biochem. Biophys. Res. Commun. 59, 108.
- 2 Kessler, C. & Manta V. (1990) Gene 92, 1-248.
- 3 Rebase The Restriction Enzyme Database http://rebase.neb.com

## **Ordering Information**

Product	Application	Packsize	Cat. No.
Restriction Enzymes	DNA restriction digestion	Please refer to website	
T4 DNA Ligase	Ligation of sticky- and blunt- ended DNA fragments.		10 481 220 001 10 716 359 001
SuRE/Cut Buffer Set for Restriction Enzymes	Incubation buffers A, B, L, M and H for restriction enzymes	1 ml each (10× conc. solutions)	11 082 035 001
SuRE/Cut Buffer A	Restriction enzyme incubation	5 × 1 ml (10× conc. solution)	11 417 959 001
SuRE/Cut Buffer B	Restriction enzyme incubation	$5 \times 1$ ml ( $10 \times$ conc. solution)	11 417 967 001
SuRE/Cut Buffer H	Restriction enzyme incubation	$5 \times 1$ ml ( $10 \times$ conc. solution)	11 417 991 001
SuRE/Cut Buffer L	Restriction enzyme incubation	5 × 1 ml (10× conc. solution)	11 417 975 001
SuRE/Cut Buffer M	Restriction enzyme incubation	$5 \times 1$ ml ( $10 \times$ conc. solution)	11 417 983 001
Water, PCR Grade	Specially purified, double-distilled, deionized, and	100 ml (4 vials of 25 ml) 25 ml	03 315 843 001 03 315 932 001
	autoclaved	(25 vials of 1 ml) 25 ml (1 vial of 25 ml)	03 315 959 001

Changes to	
previous versio	n

Editorial changes

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## **Commonly used bacterial strains**

Strain	Genotype
BL21	<i>E. coli B F <sup>-</sup> dcm ompT hsdS(r<sub>B</sub>- m<sub>B</sub>-) gal</i> (Studier, F.W. <i>et al</i> (1986) <i>J. Mol. Biol.</i> , <b>189</b> , 113.)
C600 <sup>e</sup>	supE44 hsdR2 thi-1 thr-1 leuB6 lacY1 tonA21; (Hanahan, D. (1983) J. Mol. Biol. <b>166</b> , 557.)
DH5α	supE44 Δ(lacU169 (φ80d/acZΔM15) hsdR17 recA1 endA1 gyrA96 thi-1 relA1; (Hanahan, D. (1983) J. Mol. Biol. <b>166</b> , 557.)
HB101	supE44 hsdS20 recA13 ara-14 proA2 lacY1 galK2 rpsL20 xyl-5 mtl-1; (Hanahan, D., (1983) J. Mol. Biol. <b>166</b> , 557.)
JM108	recA1 supE44 endA1 hsdR17 gyrA96 relA1 thi $\Delta$ (lac-proAB); (Yanisch- Perron, C. et al., (1985) Gene <b>33</b> , 103.)
JM109	recA1 supE44 endA1 hsdR17 gyrA96 relA1 thi $\Delta$ (lac-proAB) F[traD36proAB <sup>+</sup> , lacl <sup>q</sup> lacZ $\Delta$ M15]; (Yanisch- Perron, C. et al., (1985) Gene <b>33</b> , 103.)
JM110	rpsL (Str <sup>f</sup> ) thr leu thi-I lacY galK galT ara tonA tsx dam dcm supE44 $\Delta$ (lac-proAB) F[traD36proAB <sup>+</sup> , lacf <sup>f</sup> lacZ $\Delta$ M15]; (Yanisch- Perron, C. et al., (1985) Gene <b>33</b> , 103.)
K802	supE hsdR gal metB; (Raleigh, E. et al., (1986) Proc.Natl. Acad.Sci USA, 83, 9070.; Wood, W.B. (1966) J. Mol. Biol., <b>16</b> , 118.)
SURE <sup>r</sup>	recB recJ sbc C201 uvrC umuC::Tn5(kan') lac , Δ(hsdRMS) endA1 gyrA96 thi relA1 supE44 F'[proAB <sup>+</sup> lacI <sup>q</sup> lacZΔM15 Tn10 (tet'); (Greener, A. (1990) Stratagies, <b>3</b> , 5.)
TG1	supE hsd Δ5 thi Δ(lac-proAB) F[traD36proAB <sup>+</sup> , lacl <sup>q</sup> lacZΔM15]; (Gibson, T.J. (1984) PhD Theses. Cambridge University, U.K.)
XL1-Blue <sup>r</sup>	supE44 hsdR17 recA1 endA1 gyrA46 thi relA1 lac F'[proAB <sup>+</sup> , lacl <sup>q</sup> lacZ $\Delta$ M15 Tn10 ( $tet^0$ ]; (Bullock et al., (1987) BioTechniques, 5, 376.)

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