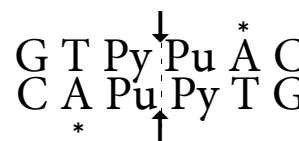


For life science research only.
Not for use in diagnostic procedures.

Restriction Endonuclease Hind II

From *Haemophilus influenzae* Rd com-10



Cat. No. 10 656 305 001 2500 units (3–10 U/ μ l)

Version 23

Content version: June 2017

Store at –15 to –25°C

Stability/Storage The undiluted enzyme solution is stable when stored at –15 to –25°C until the control date printed on the label. Do not store below –25°C to avoid freezing.

Sequence specificity *Hind* II recognizes the sequence GTPy/PuAC and generates fragments with blunt ends (1).

Compatible ends *Hind* II generates fragments with blunt ends and is compatible to any other blunt end.

Isoschizomers *Hind* II is an isoschizomer to *Hinc* II.

Methylation sensitivity *Hind* II is inhibited by 6-methyladenine as indicated (*). Furthermore *Hind* II is inhibited by 5-hydroxymethylcytosine at the 3'-C residue of the recognition sequence.

Storage buffer 20 mM Tris-HCl, 50 mM NaCl, 0.1 mM EDTA, 15 mM Dithioerythritol, 0.05% Polydocanol, 50% Glycerol (v/v), pH approx. 7.5 (at 4°C).

Suppl. Incubation buffer (10x) 100 mM Tris-HCl, 500 mM NaCl, 100 mM MgCl₂, 10 mM Dithioerythritol, pH 7.5 (at 37°C), (Δ SuRE/Cut Buffer **M**).

Activity in SuRE/Cut Buffer System Bold face printed buffer indicates the recommended buffer for optimal activity:

A	B	L	M	H
100%	100%	25–50%	100%	50–75%

Incubation temperature **37°C**

Unit definition One Unit is the enzyme activity that completely cleaves 1 μ g λ DNA in 1 h at **37°C** in the SuRE/Cut Buffer **M** in a total volume of 25 μ l.

Typical experiment

Component	Final concentration
DNA	1 μ g
10 \times SuRE/Cut Buffer M	2.5 μ l
Sterile redist. water	Up to a total volume of 25 μ l
Restriction enzyme	1 unit

Incubate at **37°C** for 1 h.

Heat Inactivation *Hind* II can be heat inactivated by 15 min incubation at 65°C.

Number of cleavage sites on different DNAs (2):

λ	Ad2	SV40	Φ X174	M13mp7	pBR322	pBR328	pUC18
35	25	7	13	2	2	2	1

Activity in PCR buffer

Relative activity in PCR mix (Taq DNA Polymerase buffer) is **100%**. The PCR mix contained λ target DNA, primers, 10 mM Tris-HCl (pH 8.3, 20°C), 50 mM KCl, 1.5 mM MgCl₂, 200 μ M dNTPs, 2.5 U Taq DNA polymerase. The mix was subjected to 25 amplification cycles.

Ligation and recutting assay

Hind II fragments obtained by complete digestion of 1 μ g λ DNA are ligated with 1 U T4-DNA ligase (Cat. No. 10 481 220 001) in a volume of 10 ml by incubation for 16 h at 25°C in 66 mM Tris-HCl, 5 mM MgCl₂, 5 mM dithioerythritol, 1 mM ATP, pH 7.5 (at 20°C) resulting in >70% recovery of 1 μ g λ DNA \times *Hind* II fragments. Subsequent re-cutting with *Hind* II yields >95% of the typical pattern of λ DNA \times *Hind* II fragments

Troubleshooting

A critical component is the DNA substrate. Many compounds used in the isolation of DNA e.g. phenol, chloroform, EtOH, SDS, high levels of NaCl, metals (e.g. Hg²⁺, Mn²⁺) inhibit or alter recognition specificity of many restriction enzymes. Such compounds should be removed by EtOH precipitation followed by drying, before the DNA is added to the restriction digest reaction.

Quality control

Lot-specific certificates of analysis are available at www.lifescience.roche.com/certificates.

Absence of unspecific endonuclease activities

1 μ g λ DNA or pBR322 DNA is incubated for 16 h in 50 μ l SuRE/Cut buffer M with excess of *Hind* II. The number of enzyme units which do not change the enzyme-specific pattern is stated in the certificate of analysis.

Absence of exonuclease activity

Approx. 5 μ g [³H] labeled calf thymus DNA are incubated with 3 μ l *Hind* II for 4 h at 37°C in a total volume of 100 μ l 50 mM Tris-HCl, 10 mM MgCl₂, 1 mM dithioerythritol, pH approx. 7.5. Under these conditions, no release of radioactivity is detectable, as stated in the certificate of analysis.

References

- Kelly, Jr., T. J. & Smith, H. O. (1970) *J. Mol. Biol.* **51**, 393.
- Kessler, C. & Manta, V. (1990) *Gene* **92**, 1–248.
- Rebase The Restriction Enzyme Database: <http://rebase.neb.com>

Ordering Information

Product	Application	Packsizes	Cat. No.
Restriction Enzymes	DNA restriction digestion	Please refer to website	
T4 DNA Ligase	Ligation of sticky- and blunt-ended DNA fragments.	100 U 500 units (1 U/μl)	10 481 220 001 10 716 359 001
SuRE/Cut Buffer Set for Restriction Enzymes	Incubation buffers A, B, L, M and H for restriction enzymes	1 ml each (10× conc. solutions)	11 082 035 001
SuRE/Cut Buffer A	Restriction enzyme incubation	5 × 1 ml (10× conc. solution)	11 417 959 001
SuRE/Cut Buffer B	Restriction enzyme incubation	5 × 1 ml (10× conc. solution)	11 417 967 001
SuRE/Cut Buffer H	Restriction enzyme incubation	5 × 1 ml (10× conc. solution)	11 417 991 001
SuRE/Cut Buffer L	Restriction enzyme incubation	5 × 1 ml (10× conc. solution)	11 417 975 001
SuRE/Cut Buffer M	Restriction enzyme incubation	5 × 1 ml (10× conc. solution)	11 417 983 001
Water, PCR Grade	Specially purified, double-distilled, deionized, and autoclaved	100 ml (4 vials of 25 ml)	03 315 843 001
		25 ml (25 vials of 1 ml)	03 315 932 001
		25 ml (1 vial of 25 ml)	03 315 959 001

Changes to previous version

Editorial changes

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Commonly used bacterial strains

Strain	Genotype
BL21	<i>E. coli</i> B F ⁻ <i>dcm ompT hsdS</i> (r _B - m _B -) <i>gal</i> (Studier, F.W. <i>et al</i> (1986) <i>J. Mol. Biol.</i> , 189 , 113.)
C600 ^e	<i>supE44 hsdR2 thi-1 thr-1 leuB6 lacY1 tonA21</i> ; (Hanahan, D. (1983) <i>J. Mol. Biol.</i> 166 , 557.)
DH5α	<i>supE44 Δ(lacU169 (φ80d/lacZΔM15) hsdR17 recA1 endA1 gyrA96 thi-1 relA1</i> ; (Hanahan, D. (1983) <i>J. Mol. Biol.</i> 166 , 557.)
HB101	<i>supE44 hsdS20 recA13 ara-14 proA2 lacY1 galK2 rpsL20 xyl-5 mtl-1</i> ; (Hanahan, D., (1983) <i>J. Mol. Biol.</i> 166 , 557.)
JM108	<i>recA1 supE44 endA1 hsdR17 gyrA96 relA1 thi Δ(lac-proAB)</i> ; (Yanisch-Perron, C. <i>et al.</i> , (1985) <i>Gene</i> 33 , 103.)
JM109	<i>recA1 supE44 endA1 hsdR17 gyrA96 relA1 thi Δ(lac-proAB) F[traD36proAB⁺, lac^q lacZΔM15]</i> ; (Yanisch-Perron, C. <i>et al.</i> , (1985) <i>Gene</i> 33 , 103.)
JM110	<i>rpsL (Str^r) thr leu thi-1 lacY galK galT ara tonA tsx dam dcm supE44 Δ(lac-proAB) F[traD36proAB⁺, lac^q lacZΔM15]</i> ; (Yanisch-Perron, C. <i>et al.</i> , (1985) <i>Gene</i> 33 , 103.)
K802	<i>supE hsdR gal metB</i> ; (Raleigh, E. <i>et al.</i> , (1986) <i>Proc. Natl. Acad. Sci. USA</i> , 83 , 9070.; Wood, W.B. (1966) <i>J. Mol. Biol.</i> , 16 , 118.)
SURE ^f	<i>recB recJ sbc C201 uvrC umuC::Tn5(kan^r) lac</i> , Δ(<i>hsdRMS</i>) <i>endA1 gyrA96 thi relA1 supE44 F[proAB⁺ lac^q lacZΔM15 Tn10 (tet^r)</i> ; (Greener, A. (1990) <i>Stratagies</i> , 3 , 5.)
TG1	<i>supE hsd Δ5 thi Δ(lac-proAB) F[traD36proAB⁺, lac^q lacZΔM15]</i> ; (Gibson, T.J. (1984) <i>PhD Theses. Cambridge University, U.K.</i>)
XL1-Blue ^f	<i>supE44 hsdR17 recA1 endA1 gyrA46 thi relA1 lac F[proAB⁺, lac^q lacZΔM15 Tn10 (tet^r)</i> ; (Bullock <i>et al.</i> , (1987) <i>BioTechniques</i> , 5 , 376.)

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